**Review** 



# The next Green Revolution: integrating crop architectype and physiotype

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In the middle of the last century, the Green Revolution dramatically increased crop yields and transformed global agriculture. As current food production is increasingly challenged by the demands of the growing population, climate change, and environmental degradation, a new Green Revolution is urgently needed. This Review highlights recent progress in defining the morphological ideotypes of four major crops, and proposes essential physiological traits critical for crop improvement and environmental adaptation. We introduce two concepts: the 'architectype' representing optimized morphological features, and the 'physiotype' encompassing improved physiological traits. By integrating these concepts through advanced genomic technologies and precision management practices, the next Green Revolution could potentially enhance crop yields and resource use efficiency by over 20–30%, thereby ensuring sustainable food production.

#### Challenges in enhancing crop production

The growing world population and rising demands for meat and dairy products necessitate continuous improvements in crop production for food and feed. Despite significant yield gains during the first Green Revolution in the 20th century (Box 1), recent decades have seen stagnation or even declines in yield for major crops in many regions (Figure 1). A number of factors such as climate change, soil degradation, water scarcity, suboptimal management practices, and socioeconomic constraints contribute to this plateau, and raise serious concerns about food security in the future [5]. In view of accelerating global warming and environmental degradation, improving crop yields and, at the same time, enhancing the resilience of crop plants to abiotic stresses (e.g., drought, heat, and flooding) and reducing the adverse environmental impact of agriculture, represents one of the biggest challenges that mankind faces in this century. This challenge is particularly pressing in sub-Saharan Africa and densely populated countries such as China and India. Advances in genetic and genomic technologies, and new interdisciplinary approaches present opportunities for a new Green Revolution to meet future food demands with a reduced environmental footprint.

This Review examines recent progress with improving morphological features of the **ideotypes** (see Glossary) for four key crops: rice, wheat, maize, and soybean. It summarizes the physiological traits involved in determining crop yields under changing climatic conditions, and explores strategies for developing elite ideotypes using advanced genetic and genomic methods, computational tools, and precision agronomy to achieve sustainable food security.

#### The ideotype of a crop

The term 'ideotype', introduced by C. M. Donald in 1968 during the first Green Revolution, refers to an idealized plant model designed to maximize biomass or product yield of crops (e.g., grain, oil, or other useful outputs) when cultivated under specific conditions [6]. The concept

#### Highlights

The Green Revolution must evolve to meet global food demand in times of climate change and global crisis.

The architectype represents an important breeding target to secure yields under varying climatic and environmental conditions.

Optimized physiological traits, defined as the physiotype, need to be integrated with optimized morphological traits to enhance yield potential, reduce resource input, and maximize environmental resilience.

The synergy between ideal architectype and optimal physiotype can enable a new Green Revolution driven by advancements in genomics, transgene, genomic selection, genome editing, molecular design, and precision management practices.

Emerging technologies, including highthroughput phenotyping, multi-omics approaches, machine learning, and artificial intelligence, will facilitate the discovery of gene sets for critical traits and accelerate the breeding of nextgeneration crop varieties.

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#### Box 1. The first Green Revolution

The first Green Revolution, initiated in the 1960s, profoundly enhanced global crop yields and saved over a billion people from starvation within 50 years. It was primarily driven by the breeding of semi-dwarf varieties of wheat and rice. Average yields per hectare in developing countries increased by 208% for wheat, 109% for rice, and 157% for maize between 1960 to 2000, with Southeast Asia, India, and China experiencing particularly remarkable yield surges [1].

Beyond high-yielding varieties generated by hybridization and genetic technologies (e.g., transgenic methods and markerassisted breeding), crop yield increases were supported by advancements in agronomic management, including irrigation systems, and essential agrochemicals (fertilizers, herbicides, and pesticides). Crucial genes underpinning GRV function in GA-related pathways, such as *Reduced height 1 (Rht1 or Rht-B1b)* and *Reduced height 2 (Rht2 or Rht-D1b)* in wheat [2] and *semi-dwarf 1 (sd1)* in rice [3]. These genes confer a semi-dwarf phenotype with increased tiller numbers, thus enhancing resistance to lodging, increasing the harvest index, and improving mechanical harvesting efficiency.

However, the reduced NUE of GRVs caused by their GA deficiency requires high fertilizer input, which contributes to environmental degradation and poses a significant challenge to the sustainability of agriculture [4]. In the face of increasing food demands and rising concerns about climate change and environmental degradation, holistic approaches towards optimizing the ideotype of crop varieties offer a promising strategy to ensure long-term food security while minimizing the environmental impact of agriculture.

emphasized the strategic selection of traits that enhance crop productivity and efficiency, thus going beyond the natural adaptations shaped by evolution. Ideotypes integrate morphological, physiological, and biochemical characteristics that collectively exceed the performance of existing cultivars.

A central component of ideotype design is plant architecture, which, among other features, determines the efficiency of crops to capture resources such as light, water, fertilizer at both the level of the individual and the level of the population (canopy). As our understanding of crop physiology and the available breeding technologies continue to advance, the concept of the ideotype evolves, by incorporating additional traits such as nutrient use efficiency, stress resilience, and pest resistance. In the future, new challenges driven, for example, by climate change, resource limitation, and increasing food demands will need to be considered as well. Consequently, the development of crop ideotypes is an ongoing and dynamic process, in which the growing needs for higher productivity and more sustainable agricultural practices need to be considered properly.

#### Ideal architectypes for major crops

The ideal ideotype differs between crop species. For example, tillering is a critical trait in most cereals (except maize), whereas branching is crucial in many dicots. Nonetheless, the ideotypes for all major crops share some common traits, including short stature with strong lodging



Figure 1. Average yield of four major crops from 1960 to 2022. (A) global; (B) China; (C) USA; (D) Brazil; (E) India. Yield data were obtained from FAOSTAT (https://www.fao.org/faostat/en/#data/QCL).

#### Glossary

Anthesis-silking interval (ASI): the time period between pollen shed (anthesis) and silk emergence in maize, serving as a key indicator of reproductive synchrony affecting grain yield.

**Architectype:** key morphological traits forming an ideal plant structure for yield optimization.

Awn: a bristle-like extension arising from the lemma, aiding seed dispersal and potentially enhancing

photosynthesis and drought tolerance. Crop Pan-genome Decoding

**Project:** a large-scale initiative integrating multi-omics and germplasm resources to map the genetic diversity of major crops, facilitating comprehensive gene discovery and enabling precision breeding.

Crop Phenome Project: a systematic initiative to characterize crop phenotypic traits (morphological, physiological, and biochemical) and their dynamic responses to environmental factors throughout the entire growth cycle, accelerating precision breeding for enhanced stress resilience and yield. **Culm:** the main stem of cereals (e.g., wheat), consisting of nodes and internodes, responsible for structural support and the transport of water, nutrients, and assimilates.

**Ideotype:** an idealized plant model with optimized traits for maximum dry matter production and yield under defined environmental conditions.

Nitrogen-use efficiency (NUE): the efficiency of nitrogen absorption and utilization for plant growth and yield. Panicle: the reproductive organ of rice and some other grasses, consisting of the rachis, primary and secondary branches, and spikelets, responsible for bearing grains.

**Physiotype:** the physiological characteristics of plants under specific environmental conditions.

Tassel: the male reproductive organ at the top of the maize plant, composed of a main axis, branches, spikelets, and florets, responsible for producing pollen. Tiller: a lateral shoot emerging from an axillary bud at the basal nodes of cereals and other grasses, classified as productive (bearing grain) or unproductive.

Water-use efficiency (WUE): the ratio of biomass or yield to water consumed, reflecting a plant's ability to use water efficiently.



resistance, compact and erect leaves allowing more **tillers** or branches to develop, mediumsized reproductive organs (that contribute to lodging resistance while supporting efficient reproduction), high yield, high cropping density tolerance, efficient photosynthesis, high nutrient use efficiency, and (abiotic and biotic) stress resilience.

The species-specific aspects of the ideotype address the unique biological properties and agronomic needs of the crop. The term 'ideal **architectype**' refers to the architectural component of the ideotype, and traditionally comprises key morphological traits that directly determine yield formation. Later, we summarize the desired characteristics of the architectypes of four major crops – rice, wheat, maize, and soybean – by synthesizing previous studies and identifying traits requiring further optimization to facilitate ideotype design and benefit agricultural production. Previously identified genes related to traits that are part of the ideal architectype of these crops are listed in Table 1.

#### Rice

The introduction of semi-dwarf varieties during the first Green Revolution in the 1960s and the subsequent breeding of hybrid rice in the 1970s represented milestones in worldwide rice cultivation. These advances were largely driven by the utilization of mutant alleles of the *sd1* gene and the development of a cytoplasmic male sterility (CMS) system. Professor Yuan Longping's pioneering work on hybrid rice has contributed substantially to global food security by facilitating the rapid commercialization of the new varieties. However, rice yield growth has plateaued in recent years, highlighting the need for a new leap forward. The Green Super Rice project, launched by the International Rice Research Institute (IRRI), the Chinese Academy of Agricultural Sciences, and the Bill & Melinda Gates Foundation, aims to achieve high and stable yields with reduced inputs and enhanced stress resistance. In China, the 'super hybrid rice' strategy integrates ideotype optimization with heterosis utilization, featuring tall erect-leaf canopies, lower **panicle** positions, and larger panicles to maximize yield potential [7].

Desired traits that, together, would contribute to higher vields include (i) moderate plant height (90–110 cm) with sturdy stems for lodging resistance and efficient photosynthate translocation; (ii) erect, thick, dark green flag leaves for enhanced photosynthesis; and (iii) the 'stay-green' trait characterized by delayed leaf senescence to extend the duration of photosynthetic carbon assimilation [8,9]. The balance between tiller number and spikelet number per panicle represents another critical determinant of rice yield. An optimal configuration would involve 8-10 productive tillers (in the absence of any unproductive ones), to minimize resource competition [10]; however, the ideal tiller number can vary significantly across different latitudes, as the heading date directly influences tiller formation. Compact tiller angles to enhance light interception and photosynthetic efficiency are favored for maximizing yield per unit growing area [11]. Improved grain yields can be achieved through increased panicle branching and grain numbers, as facilitated, for example, by tissue-specific manipulation of brassinosteroid (BR) contents and/or signaling [12]. Grain-filling rates can be accelerated by ensuring efficient assimilate flow from leaves to grains, which would support the formation of large, plump grains and lead to harvest indexes above 0.65 [13]. Additionally, a deep and robust root system - characterized by thick, branched roots and a high root-toshoot ratio - would support strong anchoring of plants in the soil, efficient water uptake and nutrient acquisition, and enhanced stress resilience [14]. Collectively, these features should lead to substantially increased productivity and improved sustainability of rice production (Figure 2A).

#### Wheat

A wheat ideotype was first proposed by Donald [6], and encompassed (i) short, strong stems, (ii) few, small and erect leaves, (iii) large, erect ears with **awns**, and (iv) a single **culm**. To further enhance yield and lodging resistance, an ideal plant height of 70–80 cm with optimized internode



#### Table 1. Examples of known genes involved in the control of the ideotype of rice, maize, wheat, and soybean

Crop	Trait	Gene name	Function	Refs
	Tiller	OsDHT1	dht1 mutants show dwarfism and increased tiller numbers	[81]
Rice		OsNAL11	NAL11 <sup>-923del-1552</sup> allele exhibits relatively few tillers, thick stems, and large panicles	[82]
		OsRCN22	rcn22 mutants display reduced tiller number	[83]
	Panicle/ grain	OsIPA1/SPL14	A point mutation in OsSPL14 results in enhanced lodging resistance, increased grain number per panicle and grain yield	[84]
		OsSPL13	Overexpression of OsSPL13 increases grain size, panicle length and branching, and grain number per panicle	[85]
		OsmiR396ef	mir396ef mutants show enhanced panicle number and grain size, lead to improved yield	[86]
		OsLARGE2	large2 mutants exhibit larger grains and increased grain number, with thick culms	[87]
		OsDREB1C	Overexpression of OsDREB1C increases grain weight and grain number per panicle	[13]
		OsGN1.1	NIL-GN1.1 <sup>B</sup> increases grain number per panicle and yield	[88]
		OsBRD3	Activated expression of the <i>BRD3</i> gene increases the number of secondary branches, grain number, and yield	[12]
		OsGNA	Overexpression of OsGNA increases panicle branch number, grain number per panicle, and yield	[89]
	Plant height	SD1	Mutations in SD1 reduce rice height and enhance lodging resistance	[3]
		SBI <sup>SV14</sup> allele	Overexpression of SBISV14 decreases plant height and improved lodging resistance	[90]
		OsMYB110	Inactivation of <i>MYB110</i> increases culm diameter and bending resistance, leading to enhanced lodging resistance despite increased plant height	[91]
		OsKAN1	OsKAN1 mutants present semi-dwarf and rolling leaf phenotypes	[92]
	Leaf	OsPROG1	Loss of <i>PROG1</i> gene function results in erect leaves, increased grain number per panicle and higher yield	[93]
		OsURL1	<i>url1</i> mutants display an upright growth phenotype that makes them suitable for planting under high-density conditions	[94]
		OsFLP	OsFLP mutation leads to increased leaf angle	[95]
	Tiller/spike number	TaPIL1	Overexpression of TaPIL1 reduces wheat tiller number and increases plant height	[96]
		TaCol-B5	Overexpression of TaCol-B5 increases tiller number, spikelet number, spike length, and yield	[20]
		DUO-B1	DUO-B1 knockout mutants exhibit increased spikelet number	[97]
Wheat	Grain number per spike	TaMYB72	<i>TaMYB72</i> mutation increases spike length, grain number per spike, the thousand grain weight, and yield	[98]
		TaFT-D2	TaFT-D2 mutation increases spike number, spikelet number, and yield	[99]
	Plant height	Rht1, Rht2	<i>Rht-B1b</i> ( <i>Rht1</i> ) and <i>Rht-D1b</i> ( <i>Rht2</i> ) alleles confer a semi-dwarf phenotype with reduced plant height and enhanced lodging resistance	[2]
		ZnF	ZnF deficient mutants display reduced plant height	[80]
		Rht8	Rht8 mutation decreases plant height	[100]
	Leaf	TaSPL8	The TaSPL8 knockout mutant exhibits erect leaves and increased spike number	[101]
	Root	TaSNAC8-6A	Overexpression of TaSNAC8-6A promotes lateral root development	[102]
	Plant height/ear height	ZmPIF3.3	Mutations of ZmPIF3.3 leads to a significant reduction in ear height and plant height	[103]
Maize		ZmTE1	Knockout of ZmTE1 significantly shortens internode length and leads to lower plant height	[104]
		ZmACS7	Mutations in ZmACS7 result in a reduced plant height and larger leaf angle	[105]
		ZmBELL10	ZmBELL10 regulates the number of internodes, internode spacing, and lodging resistance	[106]
		ZmCRY1b	Overexpression of ZmCRY1b significantly reduces plant height and ear height	[107]
		ZmCPK39	ZmCPK39 mutation decreases plant height	[108,109]
		ZmPUP4	Knockout of ZmPUP4 represses internode elongation and decreased plant height	[110]
	Leaf	Lac1	Lac1 knockout plants show 'smart-canopy' properties with upright leaves in the upper canopy, and relatively unfolded middle and lower leaves	[23]



Table 1. (co	Fable 1. (continued)						
Crop	Trait	Gene name	Function	Refs			
		ZmlDD14, ZmlDD15	<i>ZmIDD14</i> and <i>ZmIDD15</i> double mutations lead to a smaller leaf angle, reduced plant height, and ear height	[111]			
		bHLH30	Upon bHLH30 mutation, the upper leaf angle becomes smaller	[112]			
		TU1	Knockout of TU1 increases the number of leaves above the ear, leading to increased yield	[113]			
	Ear	KRN2	Knockout of KRN2 increased kernel numbers per ear and grain yield	[114]			
		EAD1	Overexpression of EAD1 led to greater ear length and kernel number per row	[115]			
		YIGE1	Overexpression of YIGE1 increased ear length and kernel number per row, thus enhanced grain yield	[116]			
		ZmACO2	ZmACO2 loss-of-function lines showed an increase in ear length, kernel number per row, and ear weight	[117]			
	Root	ZmPILS6	ZmPILS6 mutations cause reduced root network area and suppressed lateral root formation	[29]			
		ZmYUC2/4	The <i>Zmyuc4</i> single mutant and the <i>Zmyuc2/4</i> double mutant exhibit defects in the gravitropic response, thus leading to an enlarged brace root angle	[118]			
Soybean	Internode	DT1	A recessive $dt1$ allele hastens the termination of apical stem growth, thus decreasing both plant height and the number of nodes	[119]			
		DT2	Dt2 <sup>CR</sup> lines exhibit increased branch number, plant height, and stem node number	[30]			
		RIN1	rin1 mutants show less node number, shorter internode length, and reduced plant height	[31]			
		GmEID1	<i>Gmeid1</i> mutants presents shorter internodes, increased node number and branching, with greater yield	[120]			
		TOE4b	Overexpression of <i>TOE4b</i> decreases plant height, increases the number of nodes and branches, and increases pod number and yield	[121]			
	Petiole	GmILPA1	Knockout of GmILPA1 led to larger leaf petiole angle and shorter petiole	[122]			
	Pod and seed	GmPP2C-1	The PP2C-1 elite allele enhances seed size and weight	[123]			
		GmMFT	${\it GmMFT}$ overexpressing plants show a slight increase in seed length and a significant increase in the 100-seed weight	[124]			
		miR396	miR396 mutants have significantly larger seeds, more pods, and higher yield	[125]			

proportions has been proposed, slightly smaller than the 90 cm standard of the first Green Revolution [15,16]. Short basal internodes with larger diameters, combined with gradually increasing lengths and decreasing diameters toward the apex, would improve plant stability and biomass translocation efficiency [17]. Strengthening stem structure is critical for supporting increased grain biomass, while maintaining resistance to lodging.

The ideal wheat canopy exhibits a top-down gradient of leaf angles, with erect upper leaves enhancing light penetration to the lower-layer canopy, while more horizontal lower leaves would maximize light interception [18]. Flag leaves should be short, wide, thick, and dark green to efficiently facilitate photosynthesis and grain yield formation. Reduced tiller numbers with fewer unproductive tillers should enhance resource allocation to spikes, supporting the development of large spikes and grains with a high harvest index exceeding 0.62 [19]. Yield improvements have historically relied on increased grain numbers without compromising grain weight, making spike weight a promising target for future breeding and genetic engineering efforts [20]. A deep, well-developed root system enhances water and nutrient uptake, supporting aboveground growth and ensuring plant resilience under stressful conditions [21] (Figure 2B).

#### Maize

Long-term breeding has shown limited potential for further yield improvements in maize by increasing the ear size of individual plants. Under the constraints of limited arable land and low

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Appropriate increase in plant height (90-110 cm) (B) Thick and sturdy culms Erect, thick and dark green leaves Delayed leaf senescence 8-10 productive tillers Compact plant architecture with appropriate tiller angle Increased panicle branching and grain numbers Large grains Harvest index > 0.65 Deco prod eutoms with thick branched rests

Deep root system with thick, branched roots High root-to-shoot ratio



Compact, tower-shaped plant architecture Reduced plant height (2.2-2.5 m) Ear-to-plant height ratio of about 0.35 Shorter intermodes below the ear Thicker stems with high bending strength Upright leaves in the upper canopy Less erect leaves in the medium canopy More horizontal leaves in the lower canopy Small, upright and less branched tassels Vigorous crown root system and deep rooting Well-developed brace roots with larger root angle



Moderate height of 70-80 cm Sturdy stems Short basal internodes with larger diameters Increased lengths and decreasing diameter of internodes from the bottom to the top Short, wide, thick, and dark green flag leaves Compactly arranged erect leaves Few or no unproductive tillers Large spikes and large grains Harvest index > 0.62 Well-developed and deep root system

Appropriate plant height (80-110 cm) Shorter internode length (3.5-4.5 cm) 16-20 internodes Strong stems Few, short and convergent branches Short petioles and upright petiole angle (30° - 40°) Tower shape with large leaves in the lower canopy, and small leaves in the middle and upper canopy 3-4 pods per internode, 3-4 seeds per pod with plump grains Vigorous root system with appropriate root nodule density

#### Trends in Biotechnology

Figure 2. Ideal architectypes for rice, wheat, maize, and soybean. (A) Rice; (B) wheat; (C) maize; (D) soybean. Favorable traits that contribute to the ideal architectype are listed for each of the four crops.

nutrient utilization efficiency, dense planting has become the most effective strategy for increasing maize yields. Consequently, the ideal maize architectype should be compact and tower shaped, with upright leaves above the ear (leaf angle: 10°–18°) and high leaf orientation values [22]. A 'smart-canopy' architecture with upright leaves in the upper canopy, less erect leaves with increased length in the middle canopy, and horizontal leaves in the lower canopy minimizes shading, enhances ventilation, and improves light transmittance [23,24]. Functionally important leaves such as the ear leaf and those immediately above and below the ear should be nearly horizontal to maximize light capture and photosynthesis, with a leaf area index above 6.0 [25]. Compact **tassels** with fewer branches would reduce competition for resources with the ear and improve biomass allocation to kernels, while short **anthesis-silking intervals (ASI)** can enhance pollination and seed development under drought or high-density conditions [26].

Dense planting triggers shade avoidance responses (SAR) that lead to exaggerated stem elongation and reduced internode diameter, which together increase lodging risk. To mitigate these risks, the ideal maize plant should exhibit a height ranging from 2.2 to 2.5 m, which is shorter than most currently grown elite varieties, with an optimal ear-to-plant height ratio of 0.35–0.40, shorter internodes below the ear, and thicker stems with high bending strength to resist lodging [27]. A 'short-stature' maize variety recently introduced by seed companies enhances lodging resistance, **nitrogen-use efficiency (NUE)**, and compatibility with mechanical harvesting [28]. Additionally, a 'steep, cheap, and deep' root system, including vigorous crown roots and welldeveloped brace roots with steep angles, provides anchorage, supports water and nutrient uptake, and further improves lodging resistance [29]. All these traits will make maize more resilient and productive under dense planting conditions (Figure 2C).

#### Soybean

In contrast to the significant yield improvements in rice and wheat during the Green Revolution, soybean yields have stagnated, necessitating the development of new approaches in soybean



breeding. As a pod crop, soybean has some unique morphological features, with leaves, inflorescences, and pods growing at each node. This architecture limits the possibilities to achieve yield improvements through plant height reduction. Semi-dwarf soybean varieties with fewer internodes, unlike rice, wheat, and maize, are not effective in conferring substantial yield increases. Instead, optimizing the relationship between plant height and internode characteristics is essential. Taller soybean plants offer more internodes for fruit set, but are prone to lodging, whereas shorter plants resist lodging but have fewer internodes. Studies suggest an optimal plant height of 80–110 cm, a short internode length of 3.5–4.5 cm, and a total number of 16–20 internodes with strong stem diameter [30,31]. To maximize yields on the available arable land, increasing planting density will be key, which makes branch architecture a critical consideration. Under high-density planting conditions, soybeans with numerous, long branches would suffer from reduced light capture and also hinder growth of neighboring plants. Therefore, the ideal soybean architectype features few, short, and convergent branches with a strong main stem [32].

Additionally, large petiole angles and long petioles can cause self-shading, thus reducing lightharvesting efficiency and making the plants unsuitable for dense planting. Therefore, a short petiole with an upright angle of 30°–40° is strongly preferred [33]. An ideal canopy structure has large leaves in the lower canopy and smaller leaves in the middle and upper canopy, forming a towershaped spatial distribution [34]. Since soybean yield per plant largely depends on pod number rather than seed weight, three or four pods per internode and four plump seeds per pod are considered ideal for yield improvement [35]. Soybean roots form nodules with symbiotic diazotrophic *Rhizobium* bacteria, enabling biological nitrogen fixation, which is critical for plant growth and development. The ideal soybean ideotype should have a vigorous root system with robust root nodules, ensuring efficient nutrient uptake and translocation as well as stress resilience (Figure 2D).

#### Plasticity under high-density planting conditions

High-density planting is a critical strategy to improve agricultural productivity. In this context, crops with high plasticity – the ability to adjust growth and development in response to varying environmental factors – are crucial for optimizing yields and ensuring stability [36]. Ideal crops should exhibit specific architectural modifications such as reduced leaf angles and tiller numbers to minimize competition for light and resources, while maintaining plant height to prevent excessive shading and resource depletion, as planting density increases.

Rice, wheat, maize, and soybean display significant plasticity in response to planting density. Rice reduces tillering under high-density conditions and adapts its root architecture to water availability – forming shallow roots in flooded conditions and deeper roots in drought-prone environments [37]. Wheat adjusts tiller formation to ensure full canopy coverage ('ridge sealing'), thus optimizing light interception and suppressing weed growth [38]. Maize suppresses tiller formation under high density, focusing resources on a single dominant ear; whereas it produces additional tillers and secondary ears at low planting densities with adequate light and radiation conditions to enhance biomass and yield potential [39]. Soybean adjusts branching and pod-bearing habits, with infinite pod-bearing favoring long growth cycles and finite pod-bearing to short growth cycles or limited growth spaces [40].

Breeding for these adaptive traits is essential to develop new high-yielding varieties capable of thriving in high-density settings, while ensuring efficient resource utilization, to meet the demands of modern agriculture.

#### **Optimal physiotype**

The concept of the ideotype is dynamic in that it needs to continuously evolve by adjusting to changing climatic and environmental conditions as well as changing agronomic practices.



While plant morphology has traditionally been the primary focus of ideotype development, there is growing recognition that physiological traits are equally crucial for maximizing crop yields, as changes in physical structure alone do not directly alter physiological processes. For instance, modifying leaf angle can improve light utilization efficiency, but does not inherently enhance photosynthetic capacity. The integration of both architectural and physiological traits, with the latter referred to as the 'optimal **physiotype**' (i.e., the sum of the physiological characteristics of plants under specific environmental conditions), holds significant potential to maximize yields across diverse environments and resource availability constraints. An excellent example of such an approach is the release of IR8, the first semi-dwarf rice variety development by IRRI in 1966. IR8 featured a dwarf, compact morphology combined with enhanced physiological traits (e.g., high photosynthetic efficiency and robust grain-filling capacity). These combined features were responsible for IR8 conferring substantial yield increases, with grain yields being up to 2–3 times higher than those of traditional rice varieties [41]. Thus, developing crops with both ideal architectypes and optimal physiotypes is central to advancing the next Green Revolution.

#### High photosynthetic efficiency

As the green engine of life on Earth, photosynthesis converts the energy of the sunlight into chemical energy, thus enabling biomass production in nearly all ecosystems. However, the low energy conversion efficiency of photosynthesis (<1%) severely constrains crop yield improvement [42]. Climate change and, in particular, rising temperatures and  $CO_2$  levels, further complicate efforts to optimize photosynthesis in crops through breeding efforts.

To address these challenges, several strategies have been explored to improve photosynthesis and boost crop productivity. One approach proposed is to expand the light spectrum available for photosynthesis by incorporating the capacity to utilize far-red light into the canopy, either through architectural modification of the canopy structure or by introducing far-red light-harvesting pigments [43]. Another strategy that has been pursued focuses on genetic engineering to optimize key photosynthetic pathways, for example, by accelerating nonphotochemical quenching kinetics to help crops cope with fluctuating light conditions [44]. Additionally, modifying electron transport processes within the thylakoid membrane can further improve yield potential. Other efforts aim to enhance the Calvin cycle by upregulating gene expression or improving enzyme efficiency, which can drive significant yield improvements [45]. Moreover, enhancing photosynthesis during the post-anthesis period by introducing traits like stay green and extended grain filling can maintain high photosynthetic efficiency and stabilize yields, particularly in challenging environments [8,46].

#### Efficient nutrient utilization

Nitrogen and phosphorus are essential nutrients for plant growth, and directly influence crop yields through their role in various physiological processes. However, excessive application of nitrogen and phosphorus fertilizers has led to pollution, especially eutrophication of freshwaters, and environmental degradation. In Green Revolution varieties (GRVs), NUE has often been compromised, particularly due to the accumulation of DELLA proteins, which are associated with the semi-dwarfism trait [4]. To address the challenge of improving NUE and phosphorus use efficiency (PUE) and reducing fertilizer inputs, it is essential to break the trade-off between plant growth and nutrient utilization to achieve a more sustainable food production.

Nutrient use efficiencies are complex traits involving processes such as uptake, transport, assimilation, and remobilization. Enhancing these processes could be achieved, for example, by modifying root architecture to improve nutrient uptake, including increasing root density, surface area, and plasticity [47]. Optimizing nutrient transport within the plant is also a key aspect, with current



efforts being focused on modifying the expression of nutrient transporters, such as NRTs for nitrate, AMTs for ammonium, and PHTs for phosphate [48,49]. Additionally, regulating metabolic enzymes like nitrate reductase and glutamine synthetase, and/or regulators of nutrient signaling (e.g., PHR1, SPX) can enhance assimilation efficiency [50,51]. Improving nitrogen remobilization to developing grains and enhancing phosphorus recycling from senescing organs also represent promising approaches to optimize nutrient allocation [52,53]. The identification of critical transcription factors such as the NGR5–DELLA–GRF4, OsTCP19–DLT, and PHR1–SPX modules [54], and the optimization of the carbon–nitrogen balance by modifying the associated metabolism [55], holds potential for improving NUE and PUE. In summary, by closing knowledge gaps and engineering factors involved in nutrient acquisition and remobilization, it should be possible to significantly enhance crop resource use efficiency.

#### Stress resilience

Extreme weather events, such as drought periods, floods, and heat waves, are becoming more frequent and intense due to climate change, and pose significant threats to global crop production. Enhancing crop resilience to these stresses will require the translation of molecular insights gained in model plants to staple crops and equip them with the capacity to withstand multiple stresses under field conditions. Feasible strategies include the use of advanced breeding techniques (including genome editing and transgenic methods) as well as improved management practices and the use of microbial consortia to enhance stress tolerance [56].

A resilient root system plays a key role in maximizing water capture and also enhances tolerance to adverse conditions. Optimizing root architecture and metabolic efficiency can help crops better adapt to extreme conditions [14,57]. Modifying stomatal traits, such as size, density, aperture, and movement dynamics, can optimize the balance between photosynthesis and stress tolerance, especially under water-limited conditions [58,59]. The regulation of stress-responsive hormones [e.g., abscisic acid (ABA), gibberellin (GA)] and stress-associated signaling pathways (e.g., calcium, MAPK, TOR, and SnRK signaling) offers another feasible strategy to coordinate growth with stress tolerance [60–62]. Improving resistance to extreme temperature conditions will be particularly important for crops in tropical and subtropical regions, where heat stress is a growing concern [63].

In addition to genetic improvements, agronomic management practices and soil microbiome engineering can also play important roles in enhancing stress resilience. Diversified cropping systems, including intercropping and crop rotation, that are tailored to local environmental conditions can promote root growth, optimize nutrient distribution, and enhance soil microbiome diversity, all of which are important factors that contribute to sustainable and environmentally friendly agricultural productivity [64]. For example, high-yielding rice cultivars have been shown to reduce  $CH_4$  emissions from paddy soils [65]. Advanced technologies such as drones equipped with multispectral cameras are increasingly used for precise crop monitoring, enabling better field management and faster reaction to the changing needs of the crop in response to changing environmental conditions.

#### Coordination of physiological processes

Plants operate as integrated systems, where the coordination of multiple physiological processes, including photosynthesis, nutrient utilization, and stress responses, is essential for optimal growth and yield. These processes are highly interconnected, with improvements in one of them often impacting the others. For example, photosynthesis and nitrogen metabolism are tightly linked, as amino acid synthesis requires carbon skeletons derived from photosynthetic CO<sub>2</sub> fixation. Proteins involved in photosynthesis, such as the primary CO<sub>2</sub>-fixing enzyme



Rubisco, not only catalyze carbon fixation, but due to their sheer abundance, also serve as major nitrogen reservoirs. Enhancing the efficiency of photosynthetic carbon fixation through genetic engineering and/or exploration of its natural diversity remains a high priority for improving photosynthesis and resource use efficiency, although this has proven to be a very challenging target [66,67]. Similarly, traits like stay green and extended grain filling can sustain photosynthetic activity and nutrient remobilization, and in this way, contribute to yield stability under stress conditions. Our understanding of the (highly complex) regulation of photosynthesis and nitrogen utilization is still incomplete, and the elucidation of the regulatory networks underlying the coordination of carbon and nitrogen metabolism will be essential to facilitate new engineering approaches [68].

The **water-use efficiency (WUE)** of crop plants is receiving increasing attention in breeding, as water scarcity becomes more severe in many regions of the world. Improved root architecture, dynamic stomatal regulation, and reduced nonstomatal water loss can potentially align photosynthesis with water conservation needs, while maintaining efficient nutrient uptake and stress tolerance. At the same time, energy efficiency and, in particular, the proper balancing of ATP and NADPH production with the demands of downstream metabolism support efficient photosynthetic carbon fixation, nutrient assimilation, and stress responses [69,70]. Proper temporal and spatial regulation of source–sink relations ensures effective allocation of carbon and nutrients to developing grains or storage organs, thus maximizing productivity while maintaining resilience [71,72].

Interactions of plant roots with soil microbes can also enhance nutrient uptake, affect stress resilience, and overall plant health. Incorporating beneficial microbial consortia into breeding and crop management strategies has significant potential and can complement plant-based improvements [64,73]. Also, addressing biotic stresses such as pests and diseases alongside abiotic challenges will be essential for obtaining a holistic picture that integrates all aspects that contribute to the ideal physiotype. Leveraging genomics, transcriptomics, and artificial intelligence (AI)based tools to understand and manipulate these interconnected processes will enable the development of resilient, resource-efficient crop varieties that are adapted to climate change.

#### Multidisciplinary approaches for ideotype breeding

The domestication of crops by humans has spanned >10 000 years and is still ongoing. Modern crop breeding focuses on integrating desirable traits in domesticated plants, and leveraging the genetic diversity of traditional landraces and wild relatives. The development of new powerful computational tools, including meta-analyses, machine learning, Bayesian approaches, and artificial neural networks, combined with high-throughput phenotyping, genotyping, and multiomics analyses, will enable more precise genotype–phenotype mapping. This will facilitate gene and allele mining in large germplasm collections, thus greatly accelerating the identification of optimal trait combinations [74,75]. For example, haplotype-based genome-wide association studies enable the identification of beneficial trait combinations without the need for lengthy breeding cycles, and thus can bypass the time-consuming and labor-intensive traditional breeding methods based on crosses, back-crosses and large-scale evaluation of the progeny [76]. The application of genomic prediction techniques and the rapid identification of superior haplotype alleles can significantly accelerate the pace of crop improvement in the coming decades (Figure 3).

Advanced genome editing technologies such as clustered regularly interspaced short palindromic repeats (CRISPR)-CRISPR-associated protein (Cas) (including precision gene editing techniques such as base editing and prime editing) enable targeted and more precise genomic





#### **Trends in Biotechnology**

Figure 3. Schematic representation of key factors contributing to the next Green Revolution. New crop varieties that are based on integrating the ideal architectype with the optimal physiotype form the foundation of the next Green Revolution. The optimal coordination of carbon and nitrogen metabolism, optimized source–sink relations, and an intelligently engineered canopy structure, combined with improved photosynthetic efficiency, high nutrient use efficiency, and stress resilience, will be key features of the crops of the future. Harnessing the full potential (and the synergies) of these approaches is expected to boost average crop yields and nutrient use efficiency by over 30% and 20%, respectively, making a significant contribution to global food security and the increased sustainability of agricultural production systems. Abbreviations: Al, artificial intelligence; GWAS, genome-wide association study; NUE, nutrient-use efficiency. Figure created with BioRender.com.

manipulations and are already revolutionizing crop breeding [71,77]. These new tools, combined with spatially and temporally resolved omics analyses and systems biology approaches, are providing unprecedented insights into the regulatory mechanisms that govern complex physiological processes in plants. A particularly promising strategy facilitated by these developments involves the discovering, creating, and stacking of multiple beneficial genes, a process also referred to as 'physiological pyramiding', which aims to combine desirable traits into a single crop variety. Genome editing tools and their unparalleled capacity for multiplexing will speed up this process enormously. Integration of these precision breeding approaches with innovative farming systems such as crop rotation, intercropping, and precision irrigation can enhance nutrient use efficiency and, at the same time, mitigate the effects of environmental stresses. A study in Nebraska, USA (2005–2018), concluded that agronomic management contributed 39% to yield gains, whereas



the contribution of genetic improvements was only 13% [78], underscoring the importance of integrating breeding and genetic engineering efforts with optimized management practices (Figure 3).

The ideal crop varieties of the future should require fewer resources, including water, fertilizers, pesticides, herbicides, labor, and land, while also promoting the sustainable intensification of agricultural production systems. Smart breeding strategies, using a combination of elite genes and alleles, AI-assisted genomic prediction tools, and directed evolution, together with a synthetic biology approach, show promise for the rational design of crop varieties that can meet current and future challenges in crop production (Figure 3). As ideotype breeding extends from individual plants to canopy-level cropping systems, interdisciplinary and systems-based approaches will be crucial to address the complexity of the genetic and environmental factors involved in determining crop yield and yield stability under changing environmental conditions.

#### Concluding remarks and future perspectives

Arguably, the first Green Revolution was one of the pivotal milestones in agricultural history, as it revolutionized crop productivity and modernized agronomic practices. However, as the global population approaches 10 billion, the dual challenges of dwindling natural resources and escalating environmental stresses demand innovative solutions to ensure sustainable food security. In this Review, we have highlighted the critical need to develop crop varieties with optimal physiotypes that complement ideal architectural traits as the cornerstone of a new Green Revolution. Key priorities include the coordination of carbon and nitrogen metabolism, optimization of source-sink relationships, and enhancement of both individual plant and canopy-level performance. Traits such as the ability to automatically adjust the optimal tiller or branch numbers to the growth conditions, a robust and plastic root system adaptable to varying conditions, efficient photosynthesis, high nutrient use efficiency, and enhanced stress resilience, should be prioritized and will be indispensable for future crops in the next Green Revolution. Precise control of plant hormones, such as GA, BR, and strigolactone, represents a promising avenue for optimizing plant architectype and/or physiotype, as demonstrated by many recent studies [12,23,61,79,80]. Traits enabling efficient photosynthesis and high performance under high-density planting conditions will be pivotal, as they address the primary limitations that currently hinder yield improvement in major crops, akin to the transformative impact of semi-dwarf traits during the first Green Revolution. Synergizing these advancements is projected to increase average yield and nutrient-use efficiency by over 30% and 20%, respectively (Figure 3).

To realize this vision, systems biology approaches powered by AI, machine learning, and highthroughput phenotyping, provide a robust framework for germplasm exploration and the systematic identification of genes and allelic variation underlying complex quantitative traits (Box 2). Developing a comprehensive crop gene resource bank database that integrates genotypic and phenotypic data – encompassing architectural and physiological traits – from diverse germplasms will further accelerate the discovery and deployment of beneficial traits. In this context, it is imperative to initiate the **Crop Pan-Genome Decoding Project** and the **Crop Phenome Project**. This, in turn, will facilitate the development of AI-driven smart crop breeding platforms. The integration of these advanced tools with cutting-edge genetic engineering technologies, holds immense potential to expedite the development of crop varieties with tailored traits. In combination with precision agronomic practices, these innovations may herald the dawn of a new agricultural revolution, ensuring sustainable productivity in the face of global challenges (see Outstanding questions).

#### Outstanding questions

What constitutes the ideal architectype for specific climatic zones and environmental conditions?

What are the regulatory mechanisms that link architectype and physiotype, and how do they need to be modified to maximize crop performance?

How can the integration of ideal architectype and optimal physiotype occur most efficiently in crop breeding?

What roles can emerging technologies such as artificial intelligence and synthetic biology play in enabling the next Green Revolution?



#### Box 2. The role of AI and synthetic biology in the next Green Revolution

Together with rapid advancements in computational and genomic technologies, AI and synthetic biology are transforming agriculture by addressing critical global challenges, including climate change, population growth, and resource constraints. Al-driven high-throughput phenotyping, powered by drones and imaging sensors, enables real-time plant growth and stress response monitoring. For example, LemnaTec's PhenoAlxpert automates field phenotyping, while AI-powered tools such as the ICRISAT-Microsoft's AI Sowing App and Penn State University's PlantVillage app enhance weed identification and disease detection. Field management platforms like IRRI's Rice Crop Manager, the Africa Rice Center's RiceAdvice, and Bayer's FieldView optimize planting schedules, monitor crop health, and improve irrigation strategies and yield forecasting, thus enabling data-driven decision-making to enhance resource efficiency and productivity.

Al and machine learning algorithms analyze large-scale genomic datasets to help identify key traits associated with yield improvement, stress tolerance, and pest resistance. By deciphering genotype-to-phenotype relationships, Al models can pinpoint genes controlling complex agronomic traits such as drought resilience and nutrient uptake. Al-powered breeding platforms, including Bayer's Precision Breeding platform and IRRI's Global Al-Hybrid Rice Platform, optimize parent selection and predict hybrid performance, thus significantly accelerating breeding cycles across diverse environmental conditions

Synthetic biology complements AI by engineering biological systems to enhance crop performance. Initiatives such as Realizing Increased Photosynthetic Efficiency,  $C_4$  Rice, and Gains4Crops focus on improving photosynthetic efficiency, while the engineering of root-associated microbial consortia is pursued to enhance nitrogen-use efficiency. Biofortification efforts such as Golden Rice address nutritional deficiencies, and *de novo* domestication strategies facilitate the development of novel crop varieties with customized agronomic traits. Al plays a crucial role in these efforts by identifying genome editing targets and, in this way, enabling precise modifications of biological pathways to maximize yield potential.

Despite these advancements, several challenges remain. Many agronomically important genes have yet to be fully characterized, and complex genetic interactions and pleiotropic effects complicate trait engineering. Integrating multiomics datasets with AI models requires high-quality environmental and biological data to enhance prediction accuracy. Validating AI-driven insights at the field scale presents further technical and logistical hurdles. Overcoming these challenges will strengthen the predictive power and real-world applicability of AI and synthetic biology approaches in agriculture.

Together, AI and synthetic biology will very likely drive the next Green Revolution, accelerating agricultural innovation and bolstering global food security.

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#### **Declaration of interests**

The authors declare no competing interests.

#### References

- Pingali, P.L. (2012) Green revolution: impacts, limits, and the path ahead. Proc. Natl. Acad. Sci. U. S. A. 109, 12302–12308
- Peng, J. et al. (1999) 'Green revolution' genes encode mutant gibberellin response modulators. Nature 400, 256–261
- Sasaki, A. *et al.* (2002) Green revolution: a mutant gibberellinsynthesis gene in rice. *Nature* 416, 701–702
- Li, S. et al. (2018) Modulating plant growth-metabolism coordination for sustainable agriculture. Nature 560, 595–600
- Molotoks, A. *et al.* (2021) Impacts of land use, population, and climate change on global food security. *Food Energy Secur.* 10, e261
- Donald, C.M. (1968) The breeding of crop ideotypes. *Euphytica* 17, 385–403
- Zheng, X. et al. (2024) A historical review of hybrid rice breeding. J. Integr. Plant Biol. 66, 532–545
- Shin, D. et al. (2020) Natural variations at the Stay-Green gene promoter control lifespan and yield in rice cultivars. Nat. Commun. 11, 2819
- 9. Weng, J. et al. (2025) A high yield potential ideotype for irrigated rice: rice plant types with short culms and long, upright leaves. *Field Crop Res.* 322, 109696

- Takai, T. (2024) Potential of rice tillering for sustainable food production. J. Exp. Bot. 75, 708–720
- Wang, W. et al. (2022) Molecular basis underlying rice tiller angle: current progress and future perspectives. *Mol. Plant* 15, 125–137
- Zhang, X. et al. (2024) Enhancing rice panicle branching and grain yield through tissue-specific brassinosteroid inhibition. *Science* 383, eadk8838
- Wei, S. et al. (2022) A transcriptional regulator that boosts grain yields and shortens the growth duration of rice. *Science* 377, eabi8455
- Reynoso, M.A. et al. (2022) Gene regulatory networks shape developmental plasticity of root cell types under water extremes in rice. *Dev. Cell* 57, 1177–1192 e6
- Berry, P.M. et al. (2007) Ideotype design for lodging-resistant wheat. Euphytica 154, 165–179
- Piñera-Chavez, F.J. *et al.* (2016) Avoiding lodging in irrigated spring wheat. I. Stem and root structural requirements. *Field Crop Res.* 196, 325–336
- Zhang, Y. et al. (2016) Progress in improving stem lodging resistance of Chinese wheat cultivars. Euphytica 212, 275–286

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- Kumar, P. et al. (2024) Characterization of flag leaf morphology identifies a major genomic region controlling flag leaf angle in the US winter wheat (*Triticum aestivum L.*). Theor. Appl. Genet. 137, 205
- Dong, C. et al. (2023) Tiller Number1 encodes an ankyrin repeat protein that controls tillering in bread wheat. Nat. Commun. 14, 836
- Zhang, X. et al. (2022) TaCol-B5 modifies spike architecture and enhances grain yield in wheat. Science 376, 180–183
- Li, C. et al. (2021) Recognizing the hidden half in wheat: root system attributes associated with drought tolerance. J. Exp. Bot. 72, 5117–5133
- He, L. et al. (2020) Modeling maize canopy morphology in response to increased plant density. Front. Plant Sci. 11, 533514
- Tian, J. et al. (2024) Maize smart-canopy architecture enhances yield at high densities. Nature 632, 576–584
- Yan, Y. *et al.* (2024) Photosynthetic capacity and assimilate transport of the lower canopy influence maize yield under high planting density. *Plant Physiol.* 195, 2652–2667
- Huang, S. et al. (2017) Influence of plant architecture on maize physiology and yield in the Heilonggang River valley. Crop J. 5, 52–62
- Jafari, F. *et al.* (2024) Breeding maize of ideal plant architecture for high-density planting tolerance through modulating shade avoidance response and beyond. *J. Integr. Plant Biol.* 66, 849–864
- Zhao, Y. et al. (2022) Optimizing ear-plant height ratio to improve kernel number and lodging resistance in maize (Zea mays L.). Field Crop Res. 276, 108376
- Stokstad, E. (2023) High hopes for short corn. Science 382, 364–367
- Cowling, C.L. et al. (2024) ZmPILS6 is an auxin efflux carrier required for maize root morphogenesis. Proc. Natl. Acad. Sci. U. S. A. 121, e2313216121
- Liang, Q. et al. (2022) Natural variation of *Dt2* determines branching in soybean. *Nat. Commun.* 13, 6429
- Li, S. et al. (2023) Soybean reduced internode 1 determines internode length and improves grain yield at dense planting. *Nat. Commun.* 14, 7939
- Liu, S. et al. (2020) Toward a "Green Revolution" for soybean. Mol. Plant 13, 688–697
- Chen, L. et al. (2021) Overexpression of Gm/WB14 improves high-density yield and drought tolerance of soybean through regulating plant architecture mediated by the brassinosteroid pathway. Plant Biotechnol. J. 19, 702–716
- Zhao, W. et al. (2023) Leaf shape, planting density, and nitrogen application affect soybean yield by changing direct and diffuse light distribution in the canopy. *Plant Physiol. Biochem.* 204, 108071
- Li, Y. et al. (2021) Fine mapping of a QTL locus (QNFSP07-1) and analysis of candidate genes for four-seeded pods in soybean. Mol. Breed. 41, 71
- Alseekh, S. et al. (2025) Embracing plant plasticity or robustness as a means of ensuring food security. Nat. Commun. 16, 461
- Gao, H. et al. (2019) Molecular mechanisms underlying plant architecture and its environmental plasticity in rice. Mol. Breed. 39, 167
- Zhang, N. et al. (2024) Regulation of tillering and panicle branching in rice and wheat. J. Genet. Genomics, Published online December 14, 2024 https://doi.org/10.1016/j.jgg.2024. 12.005
- Veenstra, R.L. et al. (2023) Corn yield components can be stabilized via tillering in sub-optimal plant densities. Front. Plant Sci. 13, 1047268
- 40. Xiong, S. *et al.* (2023) Regulation of soybean stem growth habit: a ten-year progress report. *Crop J.* 11, 1642–1648
- 41. Khush, G.S. *et al.*, eds (2001) *The History of Rice Breeding: IRRI's Contribution*, International Rice Research Institute
- Zhu, X.G. et al. (2010) Improving photosynthetic efficiency for greater yield. Annu. Rev. Plant Biol. 61, 235–261
- Slattery, R.A. and Ort, D.R. (2021) Perspectives on improving light distribution and light use efficiency in crop canopies. *Plant Physiol.* 185, 34–48
- Long, S.P. et al. (2022) Into the shadows and back into sunlight: photosynthesis in fluctuating light. Annu. Rev. Plant Biol. 73, 617–648

- Croce, R. et al. (2024) Perspectives on improving photosynthesis to increase crop yield. Plant Cell 36, 3944–3973
- Zhang, J. et al. (2019) Identification and characterization of a novel stay-green QTL that increases yield in maize. Plant Biotechnol. J. 17, 2272–2285
- Lyzenga, W.J. *et al.* (2023) Getting to the roots of N, P, and K uptake. *J. Exp. Bot.* 74, 1784–1805
- Hu, B. *et al.* (2023) Genetic improvement toward nitrogen-use efficiency in rice: lessons and perspectives. *Mol. Plant* 16, 64–74
- Lambers, H. (2022) Phosphorus acquisition and utilization in plants. Annu. Rev. Plant Biol. 73, 17–42
- Xie, Y. et al. (2023) Plastid-localized amino acid metabolism coordinates rice ammonium tolerance and nitrogen use efficiency. Nat. Plants 9, 1514–1529
- Gao, Z. et al. (2019) The indica nitrate reductase gene OsNR2 allele enhances rice yield potential and nitrogen use efficiency. Nat. Commun. 10, 5207
- Perchlik, M. and Tegeder, M. (2017) Improving plant nitrogen use efficiency through alteration of amino acid transport processes. *Plant Physiol.* 175, 235–247
- Han, Y. et al. (2022) Mechanisms for improving phosphorus utilization efficiency in plants. Ann. Bot. 129, 247–258
- Liu, Q. *et al.* (2022) Improving crop nitrogen use efficiency toward sustainable Green Revolution. *Annu. Rev. Plant Biol.* 73, 523–551
- Ma, X. et al. (2023) Linking glucose signaling to nitrogen utilization by the OsHXK7-ARE4 complex in rice. *Dev. Cell* 58, 1489–1501.e5
- Benitez-Alfonso, Y. et al. (2023) Enhancing climate change resilience in agricultural crops. Curr. Biol. 33, R1246–R1261
- Yu, P. et al. (2024) Seedling root system adaptation to water availability during maize domestication and global expansion. *Nat. Genet.* 56, 1245–1256
- Chen, G. *et al.* (2024) Stomatal evolution and plant adaptation to future climate. *Plant Cell Environ.* 47, 3299–3315
- 59. Lawson, T. and Matthews, J. (2020) Guard cell metabolism and stomatal function. *Annu. Rev. Plant Biol.* 71, 273–302
- Devireddy, A.R. et al. (2021) Integration of reactive oxygen species and hormone signaling during abiotic stress. Plant J. 105, 459–476
- Guo, S. et al. (2025) Fine-tuning gibberellin improves rice alkali– thermal tolerance and yield. Nature 639, 162–171
- Liu, Y. et al. (2025) Target of rapamycin (TOR): a master regulator in plant growth, development, and stress responses. Annu. Rev. Plant Biol. https://doi.org/10.1146/annurev-arplant-083123-050311
- 63. Kan, Y. et al. (2023) The molecular basis of heat stress responses in plants. *Mol. Plant* 16, 1612–1634
- Arif, I. et al. (2020) Plant microbiome engineering: expected benefits for improved crop growth and resilience. *Trends Biotechnol.* 38, 1385–1396
- Jiang, Y. et al. (2017) Higher yields and lower methane emissions with new rice cultivars. Glob. Chang. Biol. 23, 4728–4738
- Salesse-Smith, C.E. et al. (2024) Increasing Rubisco as a simple means to enhance photosynthesis and productivity now without lowering nitrogen use efficiency. *New Phytol.* 245, 951–965
- Zhou, Y. et al. (2024) Genetic engineering of RuBisCO by multiplex CRISPR editing small subunits in rice. *Plant Biotechnol.* J. 23, 731–749
- Zhao, Z. et al. (2025) Engineering nitrogen and carbon fixation for next-generation plants. Curr. Opin. Plant Biol. 85, 102699
- Kramer, D.M. and Evans, J.R. (2011) The importance of energy balance in improving photosynthetic productivity. *Plant Physiol.* 155, 70–78
- Shameer, S. et al. (2019) Leaf energy balance requires mitochondrial respiration and export of chloroplast nadph in the light. *Plant Physiol.* 180, 1947–1961
- Lou, H. et al. (2024) Engineering source-sink relations by prime editing confers heat-stress resilience in tomato and rice. Cell 188, 530–549.e20
- Rosado-Souza, L. *et al.* (2023) Understanding source-sink interactions: progress in model plants and translational research to crops. *Mol. Plant* 16, 96–121
- 73. Ge, A.H. and Wang, E. (2025) Exploring the plant microbiome: a pathway to climate-smart crops. *Cell* 188, 1469–1485

# Trends in Biotechnology

14 Trends in Biotechnology, Month 2025, Vol. xx, No. xx



- Eraslan, G. *et al.* (2019) Deep learning: new computational modelling techniques for genomics. *Nat. Rev. Genet.* 20, 389–403
- Wei, X. et al. (2024) Genomic investigation of 18,421 lines reveals the genetic architecture of rice. Science 385, eadm8762
- Bhat, J.A. et al. (2021) Features and applications of haplotypes in crop breeding. Commun. Biol. 4, 1266
- Li, B. et al. (2024) Targeted genome-modification tools and their advanced applications in crop breeding. Nat. Rev. Genet. 25, 603–622
- Rizzo, G. et al. (2022) Climate and agronomy, not genetics, underpin recent maize yield gains in favorable environments. *Proc. Natl. Acad. Sci. U. S. A.* 119, e2113629119
- 79. Hu, Q. et al. (2024) Regulatory mechanisms of strigolactone perception in rice. *Cell* 187, 7551–7567 e17
- Song, L. et al. (2023) Reducing brassinosteroid signalling enhances grain yield in semi-dwarf wheat. Nature 617, 118–124
- Liu, T. et al. (2022) Dwarf and High Tillering1 represses rice tillering through mediating the splicing of D14 pre-mRNA. Plant Cell 34, 3301–3318
- Luo, L. et al. (2023) The DnaJ domain-containing heat-shock protein NAL11 determines plant architecture by mediating globerellin homeostasis in rice (*Oryza sativa*). New Phytol. 237, 2163–2179
- Mo, T. et al. (2024) The chloroplast pentatricopeptide repeat protein RCN22 regulates tiller number in rice by affecting sugar levels via the TB1-RCN22-RbcL module. *Plant Commun.* 5, 101073
- Jiao, Y. *et al.* (2010) Regulation of *OsSPL14* by OsmiR156 defines ideal plant architecture in rice. *Nat. Genet.* 42, 541–544
  Si, L. *et al.* (2016) *OsSPL13* controls grain size in cultivated rice.
- Nat. Genet. 48, 447–456 86. Miao, C. et al. (2020) Mutations in *MIR396e* and *MIR396f*
- increase grain size and modulate shoot architecture in rice. Plant Biotechnol. J. 18, 491–501
- Huang, L. et al. (2021) The LARGE2-APO1/APO2 regulatory module controls panicle size and grain number in rice. Plant Cell 33, 1212–1228
- Zhao, H. *et al.* (2024) A QTL GN1.1, encoding FT-L1, regulates grain number and yield by modulating polar auxin transport in rice. J. Integr. Plant Biol. 66, 2158–2174
- Zhang, J. *et al.* (2025) The DENSE AND ERECT PANICLE1-GRAIN NUMBER ASSOCIATED module enhances rice yield by repressing CYTOKININ OXIDASE 2 expression. Plant Cell 37, koae309
- Liu, C. et al. (2018) Shortened Basal Internodes encodes a gibberellin 2-oxidase and contributes to lodging resistance in rice. Mol. Plant 11, 288–299
- Wang, T. et al. (2024) The transcription factor MYB110 regulates plant height, lodging resistance, and grain yield in rice. *Plant Cell* 36, 298–323
- He, Q. et al. (2024) OsKANADI1 and OsYABBY5 regulate rice plant height by targeting GIBERELLIN 2-OXIDASE6. Plant Cell 37, koae276
- Tan, L. et al. (2008) Control of a key transition from prostrate to erect growth in rice domestication. Nat. Genet. 40, 1360–1364
- Fang, J. et al. (2021) The URL1-ROC5-TPL2 transcriptional repressor complex represses the ACL1 gene to modulate leaf rolling in rice. Plant Physiol. 185, 1722–1744
- Liu, H. et al. (2024) The rice R2R3 MYB transcription factor FOUR LIPS connects brassinosteroid signaling to lignin deposition and leaf angle. *Plant Cell* 36, 4768–4785
- Zhang, L. et al. (2022) PIL transcription factors directly interact with SPLs and repress tillering/branching in plants. New Phytol. 233, 1414–1425
- Wang, Y. et al. (2022) Improving bread wheat yield through modulating an unselected AP2/ERF gene. Nat. Plants 8, 930–939
- Wu, L. et al. (2024) TaMYB72 directly activates the expression of TaFT to promote heading and enhance grain yield traits in wheat (*Triticum aestivum* L.). J. Integr. Plant Biol. 66, 1266–1269
- Liu, H. et al. (2024) A rare natural variation in TaFT-D2 underlies QTss.cas-3D associated with increased total spikelet number per spike in wheat. Crop J. 12, 1727–1734
- 100. Xiong, H. et al. (2022) Cloning and functional characterization of Rht8, a "Green Revolution" replacement gene in wheat. Mol. Plant 15, 373–376

- Liu, K.Y. et al. (2019) Wheat TaSPL8 modulates leaf angle through auxin and brassinosteroid signaling. Plant Physiol. 181, 179–194
- 102. Mao, H. et al. (2020) Regulatory changes in TaSNAC8-6A are associated with drought tolerance in wheat seedlings. Plant Biotechnol. J. 18, 1078–1092
- Wang, B. et al. (2020) Genome-wide selection and genetic improvement during modern maize breeding. Nat. Genet. 52, 565–571
- Wang, F. et al. (2022) ZmTE1 promotes plant height by regulating intercalary meristem formation and internode cell elongation in maize. *Plant Biotechnol. J.* 20, 526–537
- Li, H. et al. (2020) Maize plant architecture is regulated by the ethylene biosynthetic gene ZmACS7. Plant Physiol. 183, 1184–1199
- 106. Yu, J. et al. (2023) ZmBELL10 interacts with other ZmBELLs and recognizes specific motifs for transcriptional activation to modulate internode patterning in maize. New Phytol. 240, 577–596
- 107. Chen, S. et al. (2024) Cryptochrome 1b represses gibberellin signaling to enhance lodging resistance in maize. *Plant Physiol.* 194, 902–917
- Zhu, M. et al. (2024) The maize ZmCPK39-ZmKnox2 module regulates plant height. aBIOTECH 5, 356–361
- 109. Zhu, M. et al. (2024) The ZmCPK39-ZmDi19-ZmPR10 immune module regulates quantitative resistance to multiple foliar diseases in maize. Nat. Genet. 56, 2815–2826
- 110. Huang, Y. et al. (2024) PURINE PERMEASE 4 regulates plant height in maize. J. Genet. Genomics 52, 446–448
- 111. Liu, D. et al. (2024) Coordinated control for the auricle asymmetric development by ZmIDD14 and ZmIDD15 fine-tune the high-density planting adaption in maize. *Plant Biotechnol. J.* 22, 2675–2687
- 112. Wang, Q. et al. (2024) Histological and single-nucleus transcriptome analyses reveal the specialized functions of ligular sclerenchyma cells and key regulators of leaf angle in maize. *Mol. Plant* 17, 920–934
- 113. Li, Y. et al. (2024) MADS-box encoding gene Tunicate1 positively controls maize yield by increasing leaf number above the ear. Nat. Commun. 15, 9799
- Chen, W. et al. (2022) Convergent selection of a WD40 protein that enhances grain yield in maize and rice. Science 375, eabg7985
- 115. Pei, Y. et al. (2022) EAR APICAL DEGENERATION1 regulates maize ear development by maintaining malate supply for apical inflorescence. Plant Cell 34, 2222–2241
- Luo, Y. et al. (2022) Genetic variation in YIGE1 contributes to ear length and grain yield in maize. New Phytol. 234, 513–526
- 117. Ning, Q. et al. (2021) An ethylene biosynthesis enzyme controls quantitative variation in maize ear length and kernel yield. Nat. Commun. 12, 5832
- Zheng, Z. *et al.* (2023) Local auxin biosynthesis regulates brace root angle and lodging resistance in maize. *New Phytol.* 238, 142–154
- 119. Liu, B. et al. (2010) The soybean stem growth habit gene Dt1 is an ortholog of Arabidopsis TERMINAL FLOWER1. Plant Physiol. 153, 198–210
- 120. Qin, C. et al. (2023) GmEID1 modulates light signaling through the Evening Complex to control flowering time and yield in soybean. Proc. Natl. Acad. Sci. U. S. A. 120, e2212468120
- Li, H. et al. (2023) The AP2/ERF transcription factor TOE4b regulates photoperiodic flowering and grain yield per plant in soybean. Plant Biotechnol. J. 21, 1682–1694
- 122. Gao, J. et al. (2017) GmlLPA1, encoding an APC8-like protein, controls leaf petiole angle in soybean. Plant Physiol. 174, 1167–1176
- Lu, X. et al. (2017) A PP2C-1 allele underlying a quantitative trait locus enhances soybean 100-seed weight. Mol. Plant 10, 670–684
- 124. Cai, Z. et al. (2023) MOTHER-OF-FT-AND-TFL1 regulates the seed oil and protein content in soybean. New Phytol. 239, 905–919
- 125. Xie, H. et al. (2024) Knockout of miR396 genes increases seed size and yield in soybean. J. Integr. Plant Biol. 66, 1148–1157